

A Novel Approach toward Bacteriochlorophylls-*e* and *f*

Hitoshi Tamiaki,^{a,b*} Miki Omoda^a and Masanobu Kubo^a

^aDepartment of Bioscience and Biotechnology, Faculty of Science and Engineering,
Ritsumeikan University, Kusatsu, Shiga 525-8577, Japan; and ^bPRESTO, JST

Received 17 March 1999; accepted 20 April 1999

Abstract: Methyl bacteriopheophorbide-*f* was prepared from methyl bacteriopheophorbide-*d* with retention of the 3¹-chirality. The transformation of the methyl to the formyl group at the 7-position of the chlorin moiety will provide an alternative route for the synthesis of bacteriochlorophylls-*e* and *f*. © 1999 Elsevier Science Ltd. All rights reserved.

Bacteriochlorophyll(=BChl)-*e* is a major extramembraneous antenna pigment in anoxygenic photosynthetic brown-colored bacteria, e.g., *Chlorobium phaeovibriodes* and *phaeobacteriodes*.¹ BChl-*e* is a magnesium complex of 7-formyl chlorin and is differentiated by the 7-substituent from BChl-*c* which possesses the 7-methyl group and is a pigment in the light-harvesting antenna of green-colored bacteria (see Fig. 1). The same relationship is seen in natural pigments of higher plants between chlorophyll(=Chl)-*b* (7-CHO) and Chl-*a* (7-CH₃). Moreover, the name BChl-*f* is reserved for the compound in which the 7-methyl group of naturally occurring BChl-*d* is substituted by a 7-formyl group: BChl-*f* has not yet been found in any photosynthetic bacteria. Methyl bacteriopheophorbide-*f* (**1**) is a derivative of BChl-*f* (demetallation and transesterification) and has been prepared from methyl pyropheophorbide-*b* by hydration of the 3-vinyl group.² Here, we report the alternative synthesis of **1** from methyl bacteriopheophorbide-*d* (**2**) by transformation of the 7-methyl to the 7-formyl group.

Addition of **2** with OsO₄ in the presence of pyridine and cleavage of the resulting cyclic ester by H₂S³ gave 7,8-*cis*-diol **3** (49%) which was *ca.* a 1:1 diastereomeric mixture at the 7,8-positions (see Scheme 1). The 7,8-double bond is the most reactive in the chlorin π -chromophore because of its relatively low conjugation, but the 3¹-hydroxyl group of the adduct was also oxidized under the above conditions to afford undesired 3-acetyl-7,8-diol in 7% yield. To suppress the over-reaction, the process of the oxidation was checked by TLC and the reaction was quenched by H₂S when the third quarter of **2** was consumed. Mild dehydration⁴ of **3** by acidic treatment gave *ca.* a 1:8 isomeric mixture of primary alcohol **4** (7¹-OH) and more stable secondary alcohol **5** (8¹-OH). Flash column chromatography over silica gel with 1% MeOH and CH₂Cl₂ successfully separated the regio-isomers and pure **4** was isolated in 10% yield. Selective oxidation of the 7-hydroxymethyl group in **4** by PDC (see legend Scheme 1)

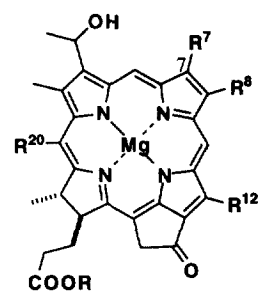


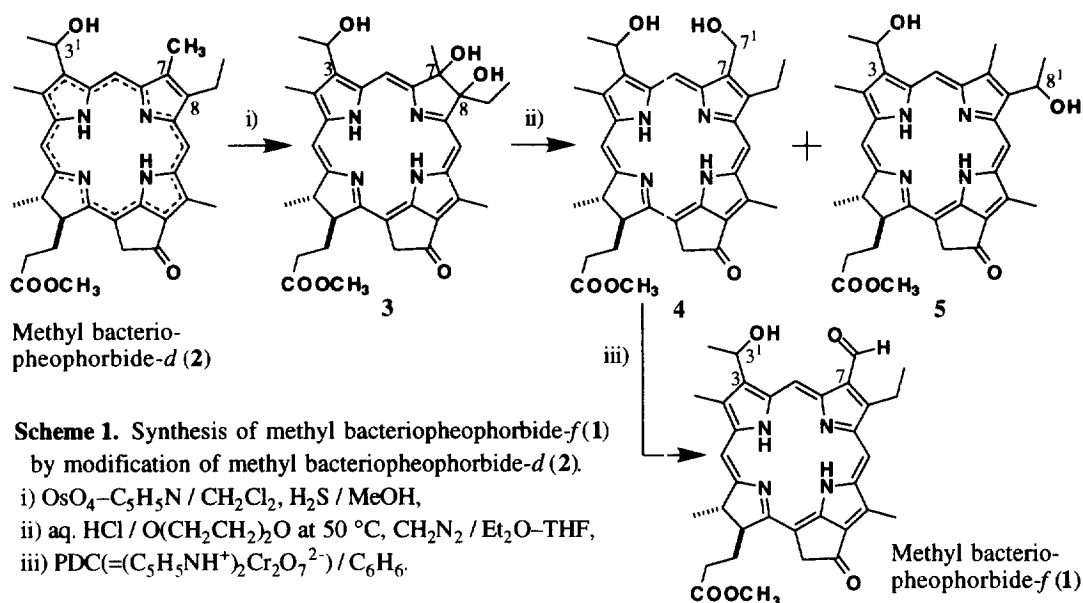
Figure 1. Photosynthetic antenna pigments.

BChl-*c*: R⁷=R²⁰=CH₃

BChl-*d*: R⁷=CH₃, R²⁰=H

BChl-*e*: R⁷=CHO, R²⁰=CH₃

BChl-*f*: R⁷=CHO, R²⁰=H



yielded **1** (79%). When a (3¹*S*)-enriched sample of **2** (*R/S*=1/9)⁵ was used as the starting material, the diastereomeric ratio of **1** produced was determined to be 1:9 from the HPLC analysis, indicating that no epimerization at the 3¹-position occurred during the transformation of the methyl to the formyl group at the 7-position. Transesterification from methyl group in **1** to a long chain (e.g., farnesyl group) followed by magnesium insertion would lead to BChl-*f* ($\text{R}^8=\text{Et}$, $\text{R}^{12}=\text{Me}$).

Molecular structures (including the stereochemistry) of various BChls-*c* and *d* separated from photosynthetic green bacteria have already been determined,¹ but many BChls-*e* have still not been confirmed in the R^8 , R^{12} and R substituents as well as the 3¹-stereochemistry. Moreover, BChl-*f* has not been observed in any antenna pigment. The present transformation of 7- CH_3 to 7- CHO with retention of the 3¹-chirality will be useful for structural assignments of BChls-*e* and *f*. The structurally determined BChls-*f* and their derivatives will be helpful for elucidation of the mechanism in biosynthesis of BChl-*e* (BChlide-*d* ($\text{R}=\text{H}$ in Fig. 1) \rightarrow BChlide-*c* \rightarrow BChlide-*e* or BChlide-*d* \rightarrow BChlide-*f* \rightarrow BChlide-*e*)⁶ as well as for the first detection in chlorophyllous pigments extracted from photosynthetic bacteria.

Acknowledgment: We thank Mr. Satoshi Sakuma, Ritsumeikan University, for his assistance in the experiments.

References

- Scheer, H. In *Chlorophylls*; Scheer, H. Ed.; CRC Press: Boca Raton FL, 1991; pp. 3–30.
- Risch, N.; Köster, B.; Schormann, A.; Siemens, T.; Brockmann, H. *Liebigs Ann. Chem.* **1988**, 343–373.
- Tamiaki, H.; Tomida, T.; Miyatake, T. *Bioorg. Med. Chem. Lett.* **1997**, 7, 1415–1418; Pandey, R. K.; Isaac, M.; MacDonald, I.; Medforth, C. J.; Senge, M. O.; Dougherty, T. J.; Smith, K. M. *J. Org. Chem.* **1997**, 62, 1463–1472.
- Inhoffen, H. H.; Jäger, P.; Mählpf, R. *Liebigs Ann. Chem.* **1971**, 749, 109–116.
- Tamiaki, H.; Takeuchi, S.; Tsudzuki, S.; Miyatake, T.; Tanikaga, R. *Tetrahedron* **1998**, 54, 6699–6718.
- Porra, R. J. *Photochem. Photobiol.* **1997**, 65, 492–516.